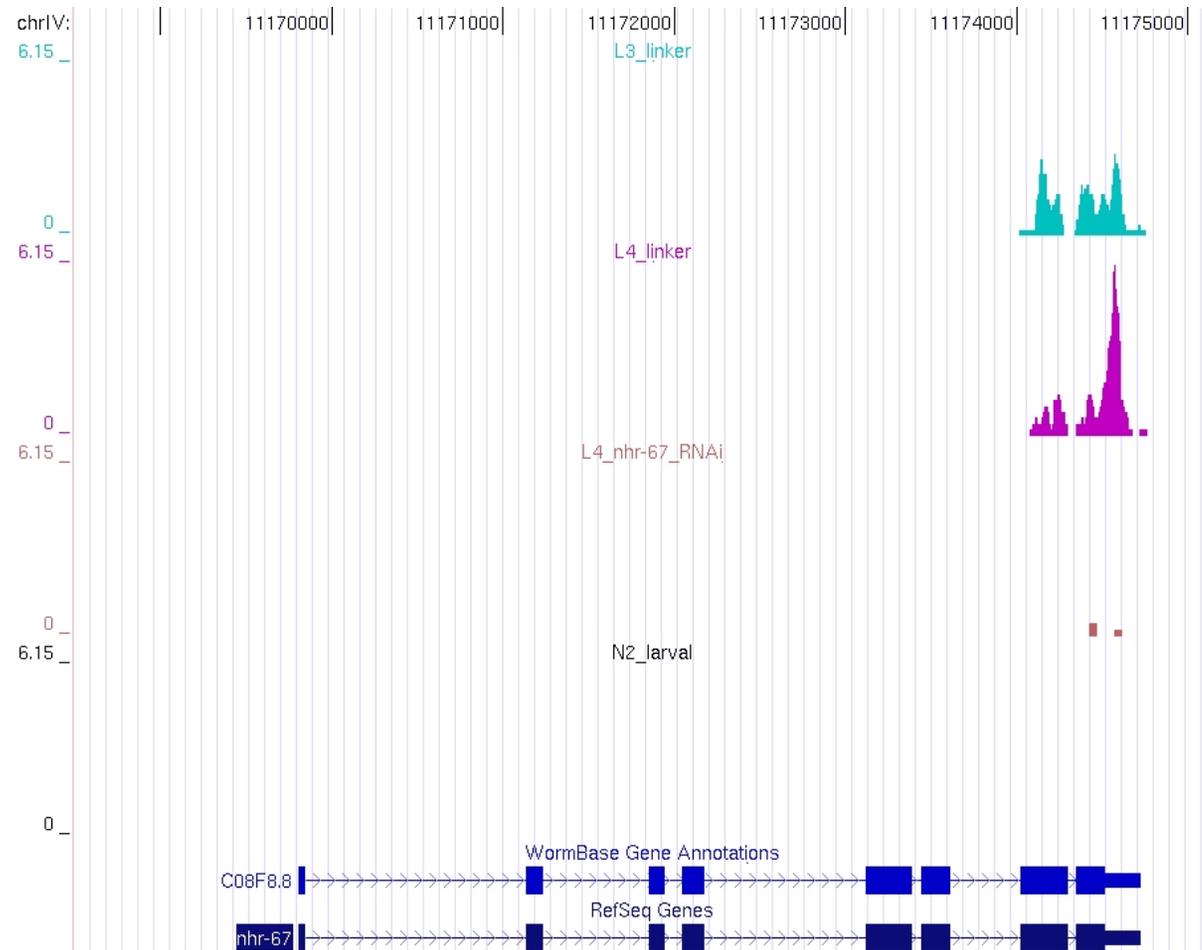


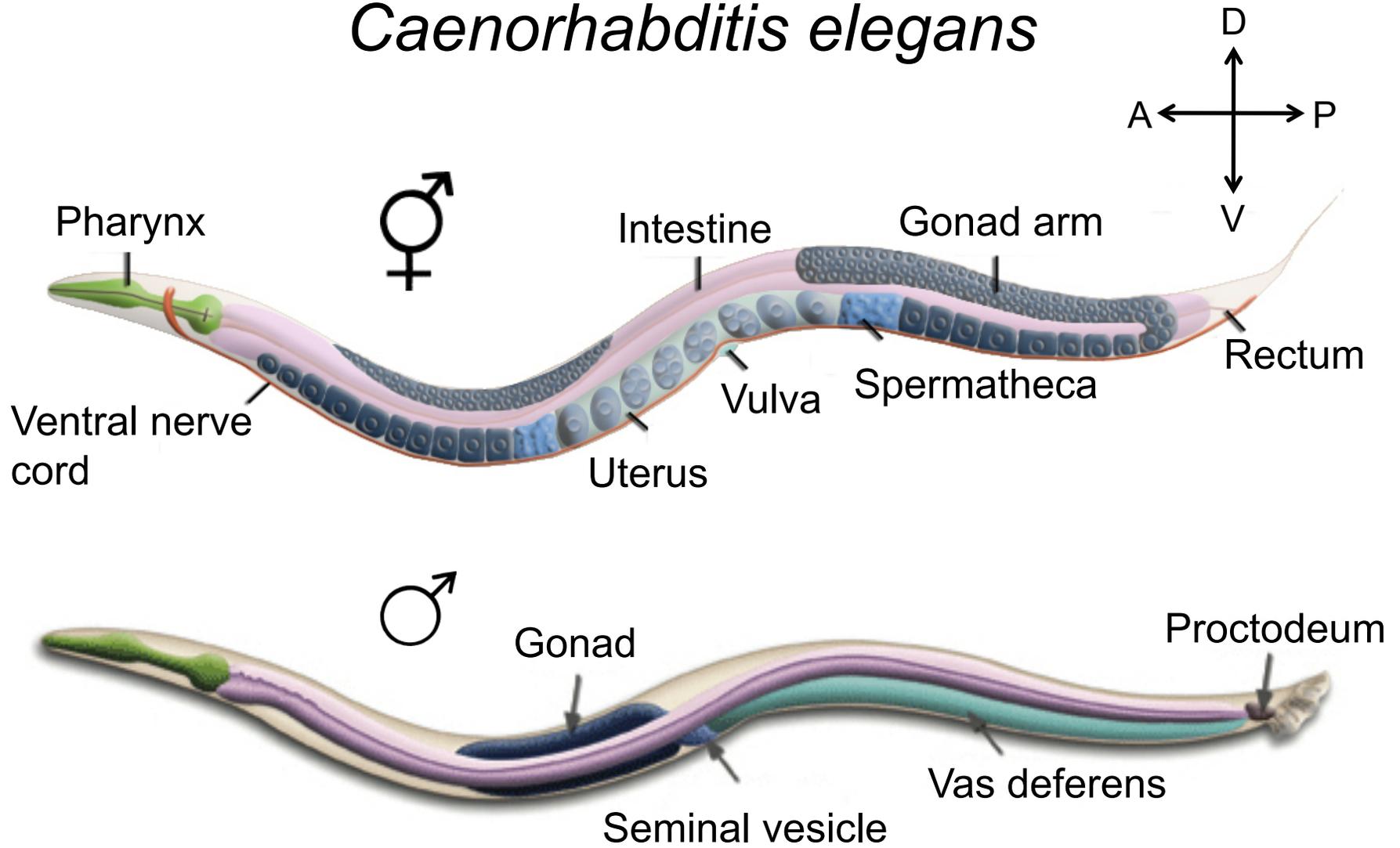
# Single-cell RNA-seq in *C. elegans*

Erich Schwarz, Cornell



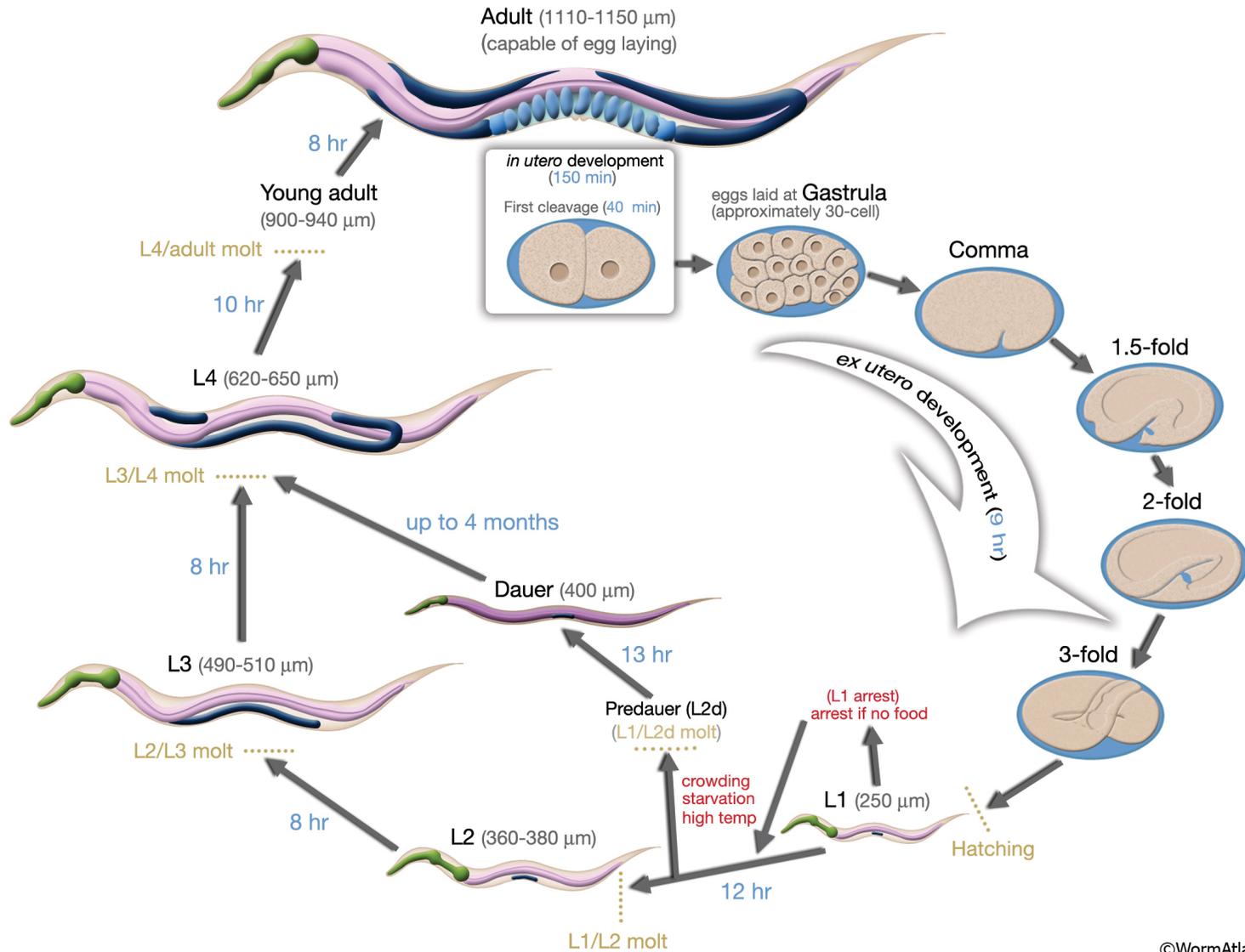
MSU NGS Course, June 2013

# *Caenorhabditis elegans*



Ref.: Hall (2013), [www.wormatlas.org](http://www.wormatlas.org).

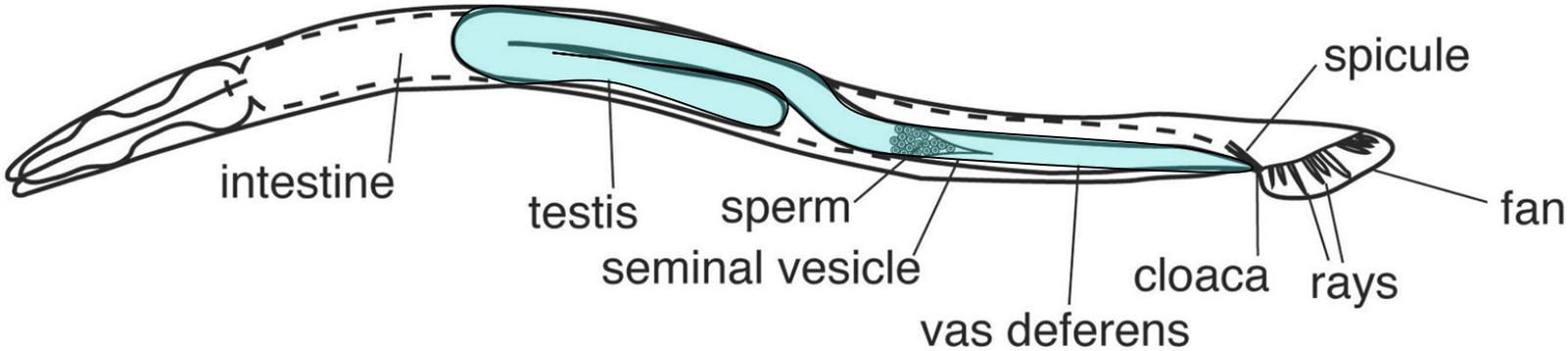
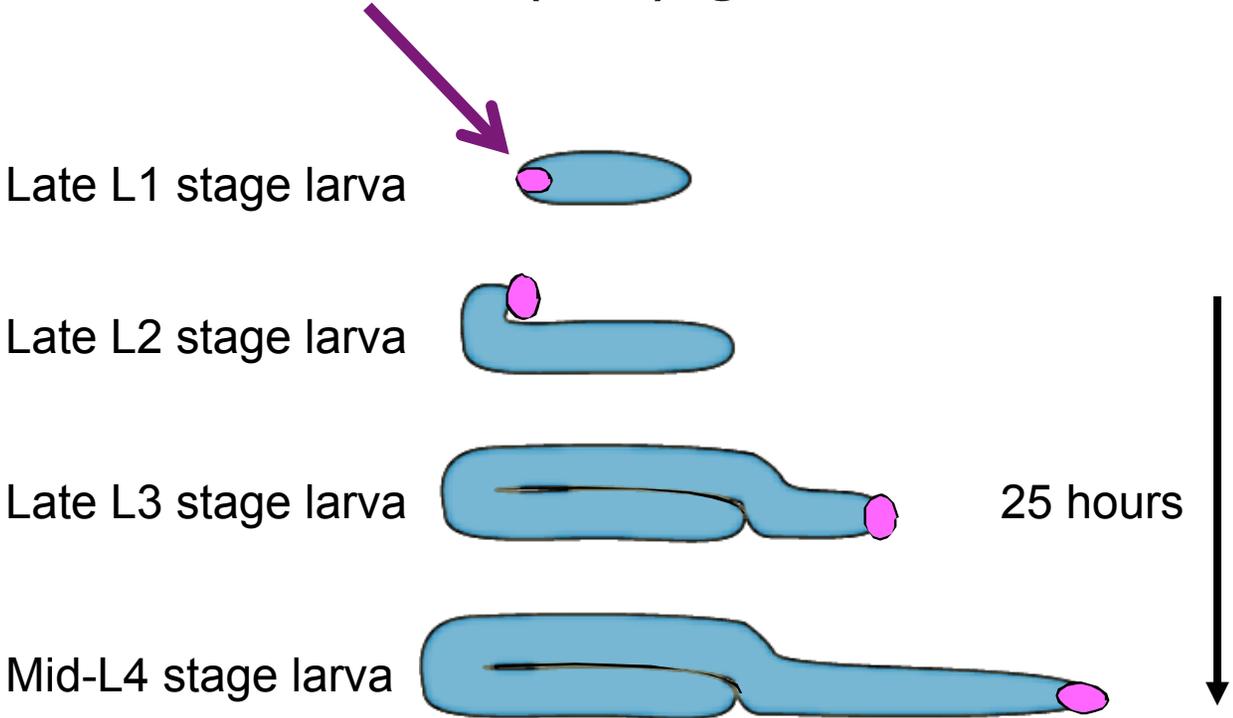
# Caenorhabditis elegans



©WormAtlas

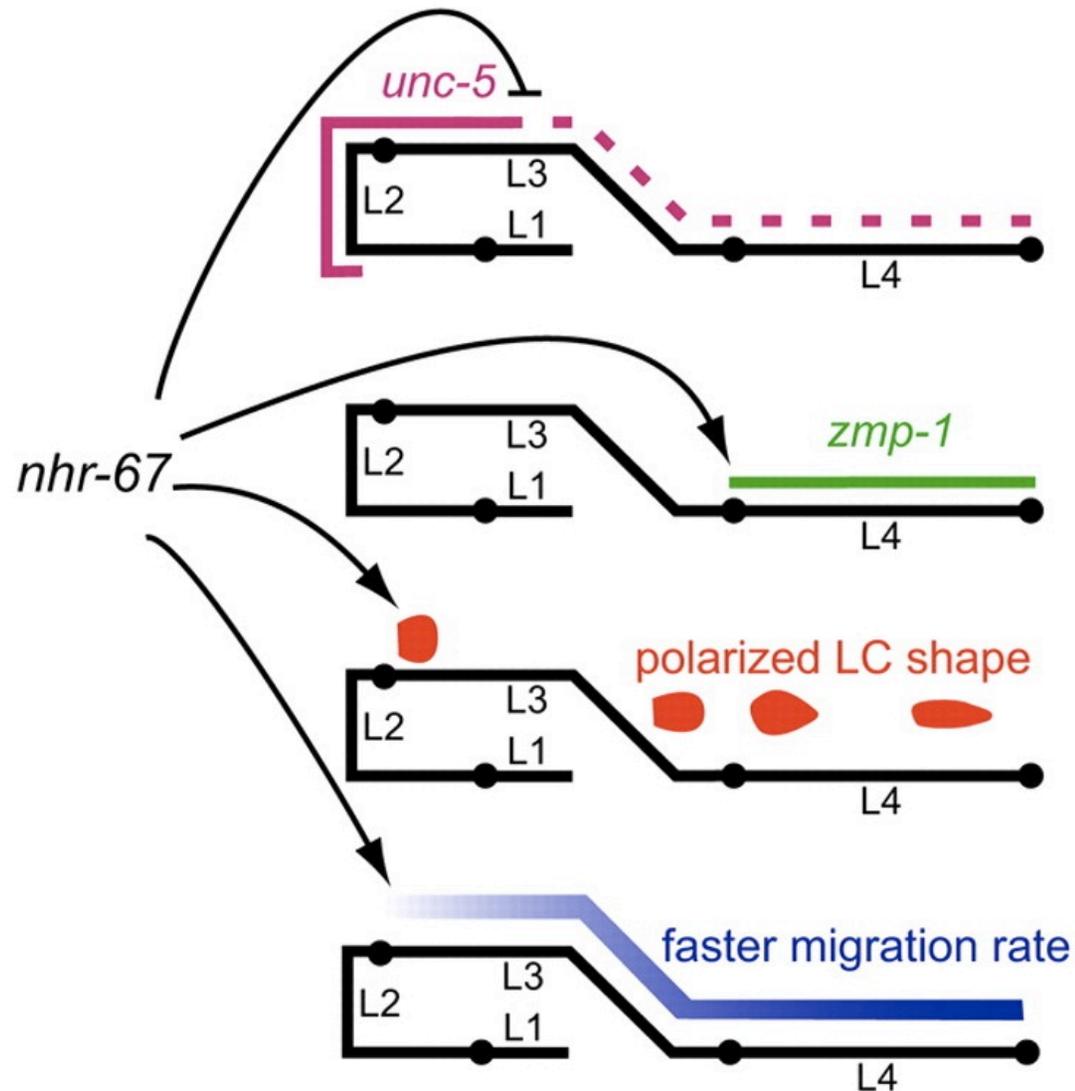
Ref.: Hall (2013), [www.wormatlas.org](http://www.wormatlas.org).

# The linker cell (LC) guides the male gonad



Ref.: Kato and Sternberg (2009), *Development*, 136, 3907-3915.

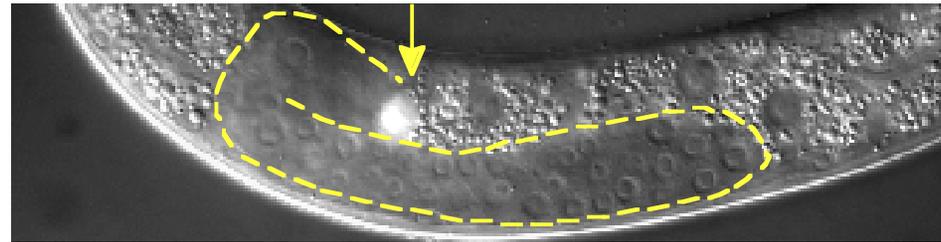
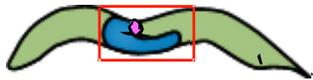
# Linker cells partially require *nhr-67* (*tailless/Tlx*)



Ref.: Kato and Sternberg (2009), *Development*, 136, 3907-3915.

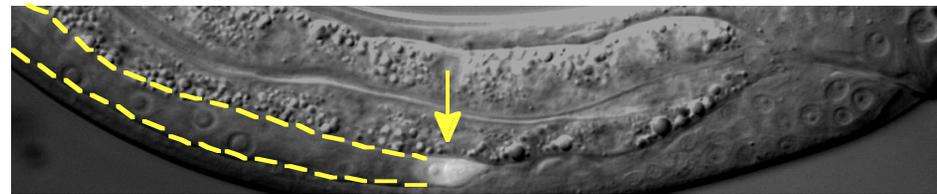
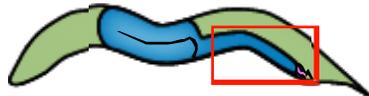
# LC samples harvested for RNA-seq

L3 stage

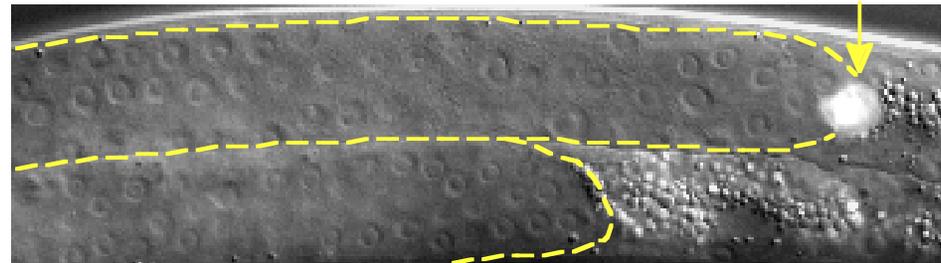


LC is labeled by YFP

L4 stage



L4, *nhr-67*(-)

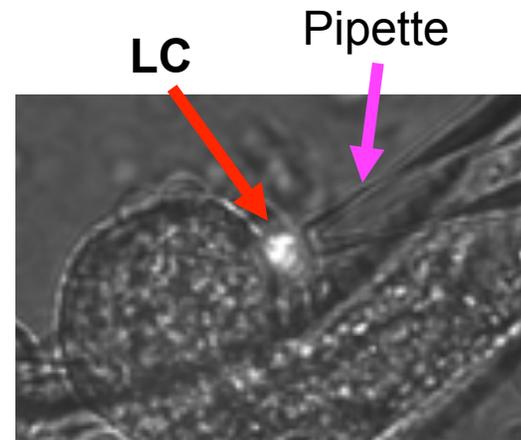
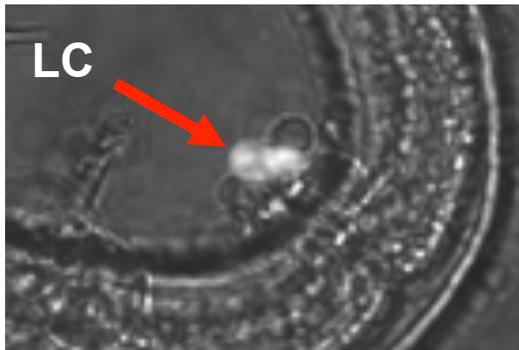
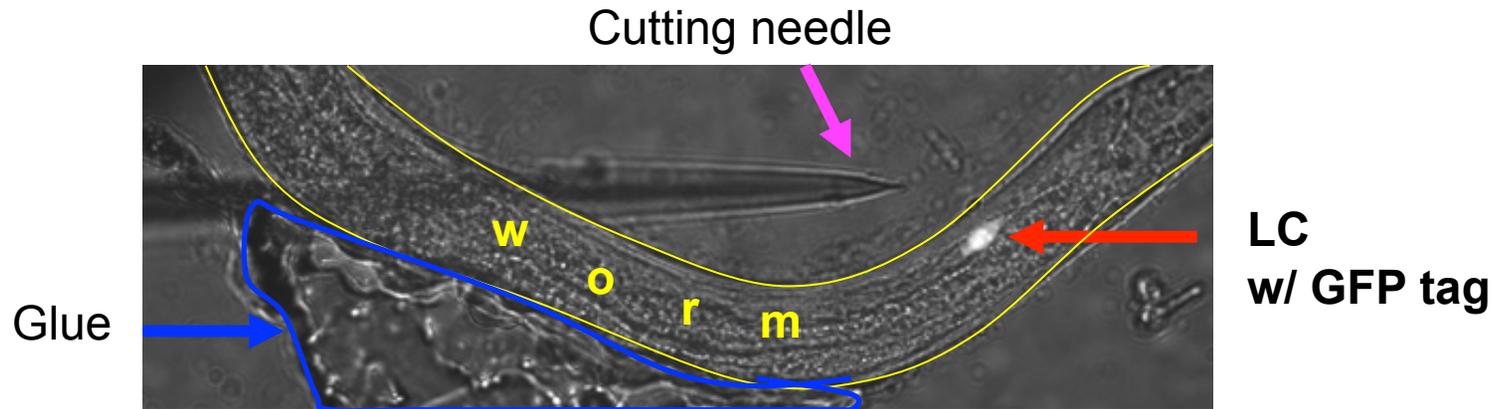


Whole L1+  
larvae



# Microdissecting individual LCs

1. Glue the worm onto an agar pad and immerse it in buffer.

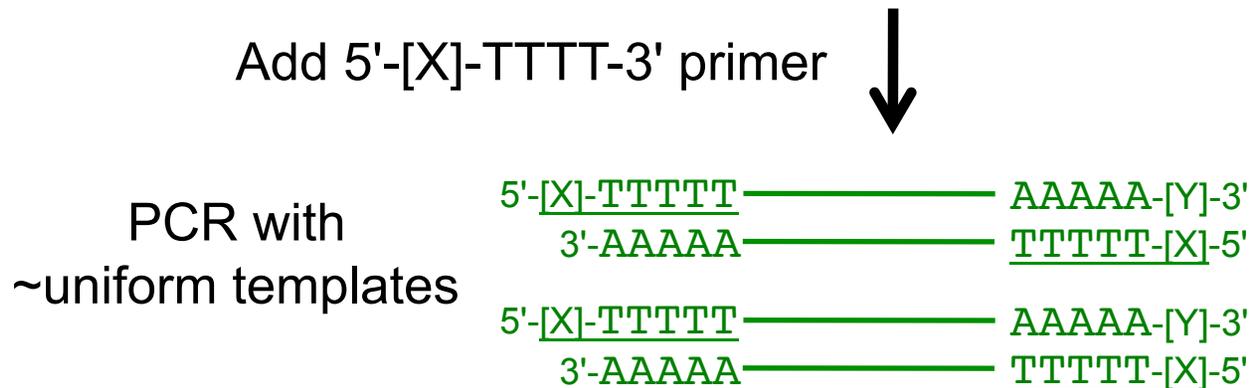


2. Cut into the worm to release the LC.

3. Suck the LC into a pipette and snap-freeze it in liquid nitrogen.

Ref.: Lockery and Goodman (1998), Meth. Enzym. 293, 201-217.

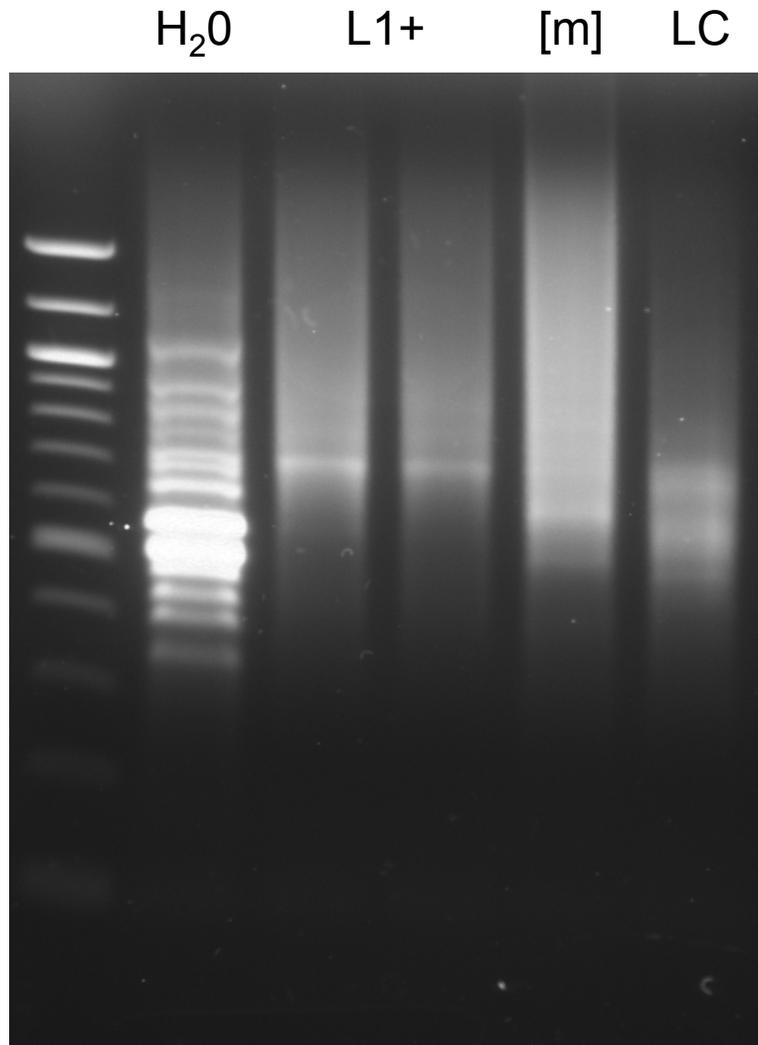
# RT-PCR of dissected cells



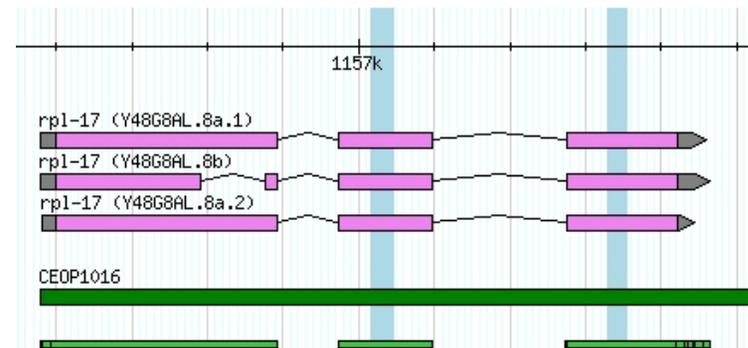
Ref.: Dulac and Axel (1995), Cell 20, 195-206.

# RT-PCR products from a single linker cell

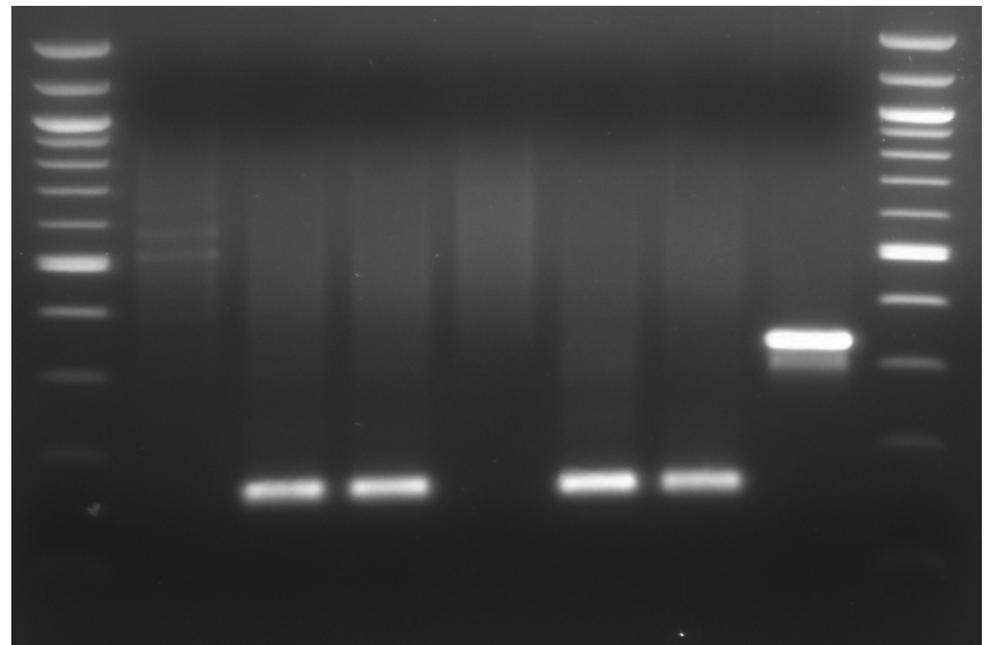
Primary RT-PCRs:



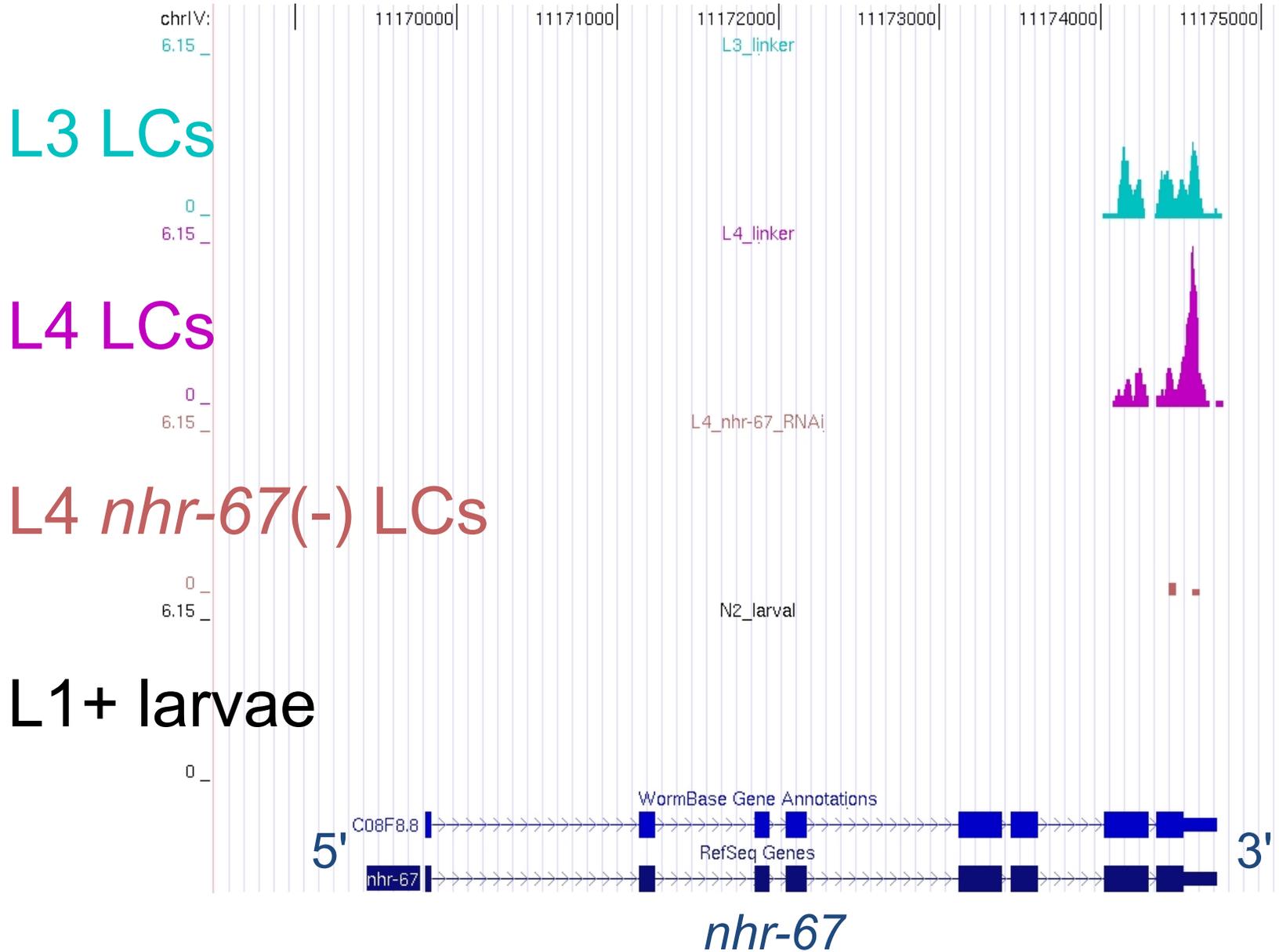
*rpl-17* test PCRs:



H<sub>2</sub>O L1+ [m] LC AFD gDNA



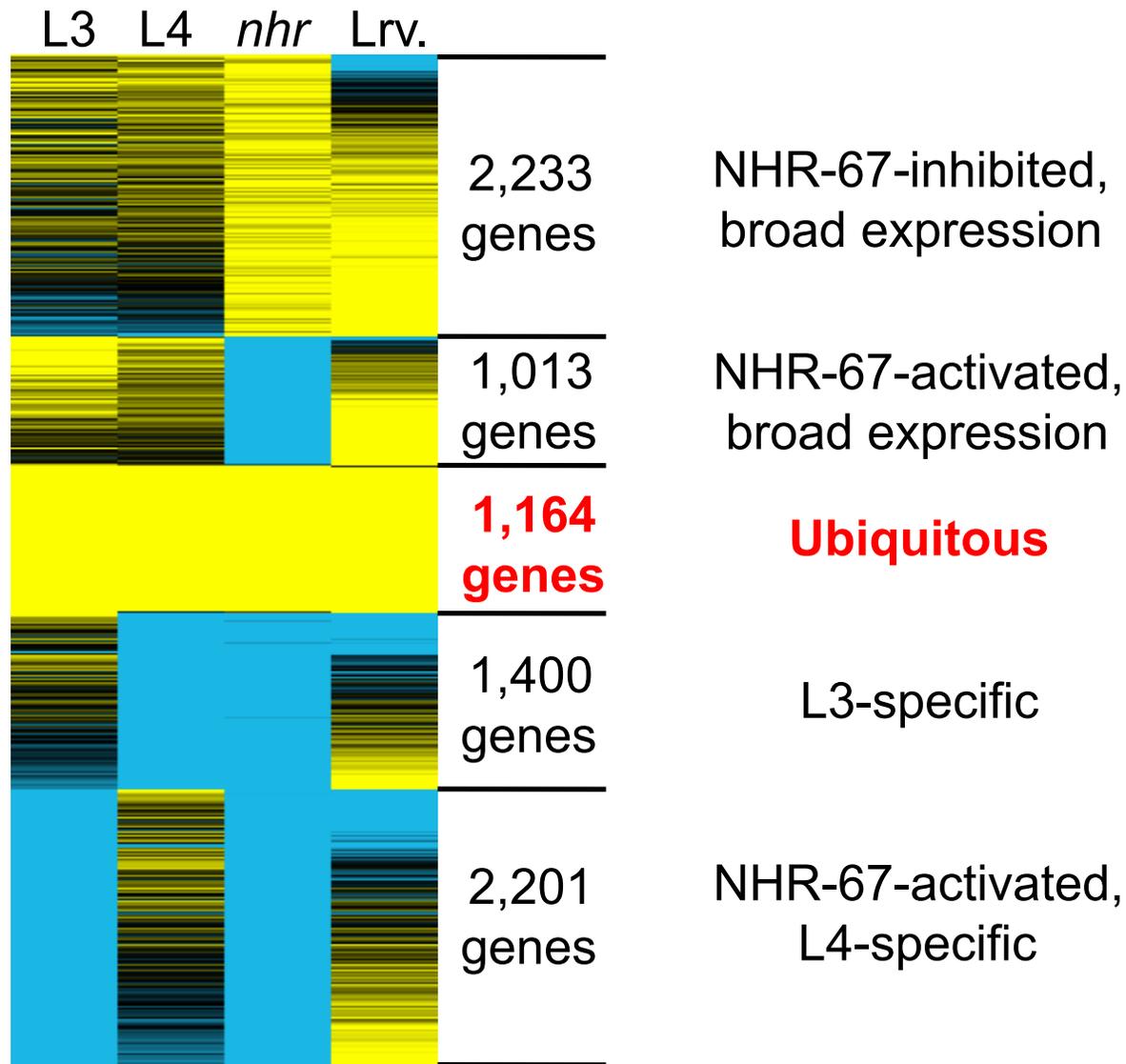
# RNA-seq of pools (five LCs or samples apiece)



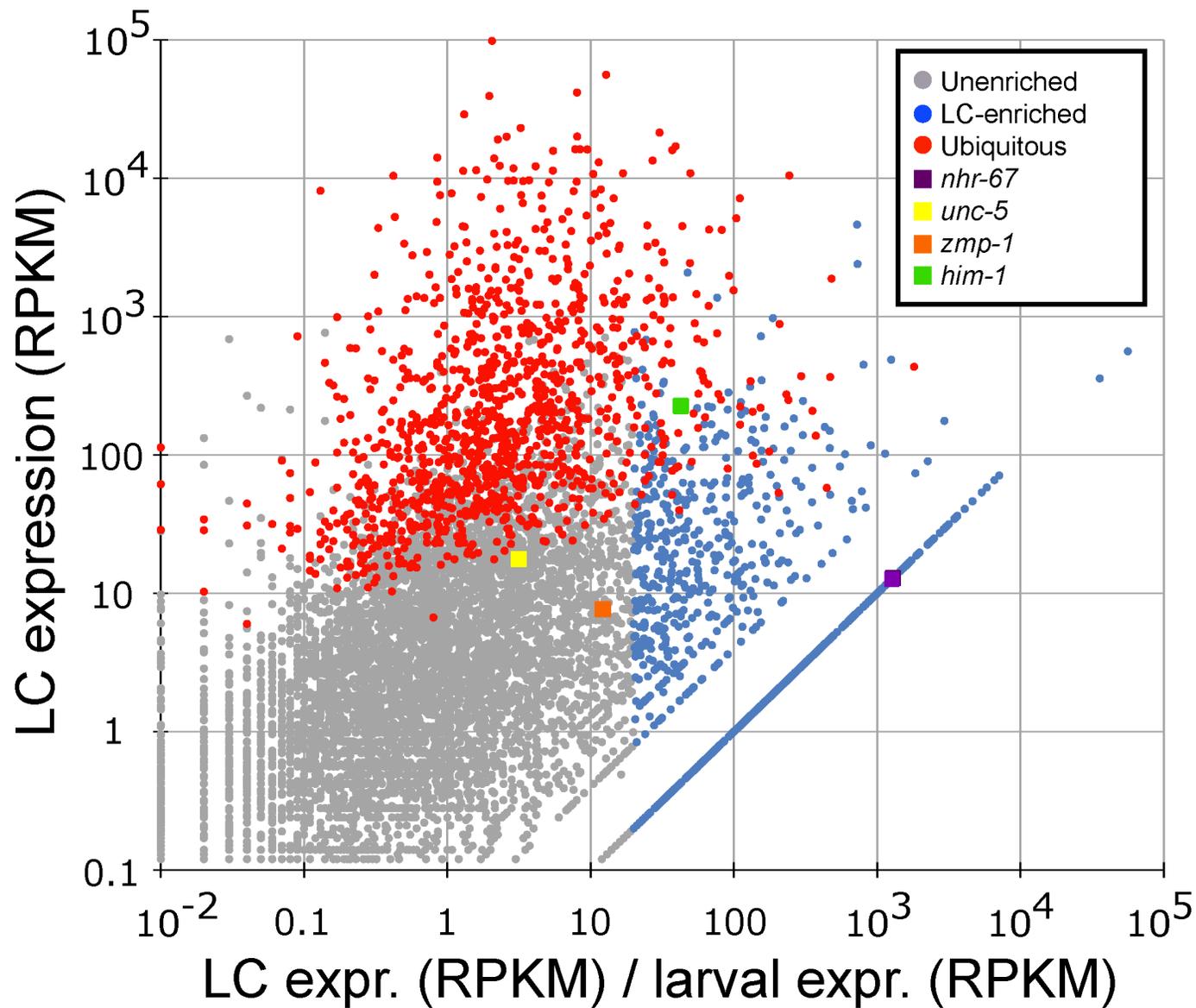
# Expressed genes

Maximum detectable	20,252
L3-stage LC	5,740
L4-stage LC	6,603
<i>nhr-67</i> (RNAi) L4 LC	5,290
L3 or L4 w.t. LC	8,011
Wild-type L1+ larvae	13,152

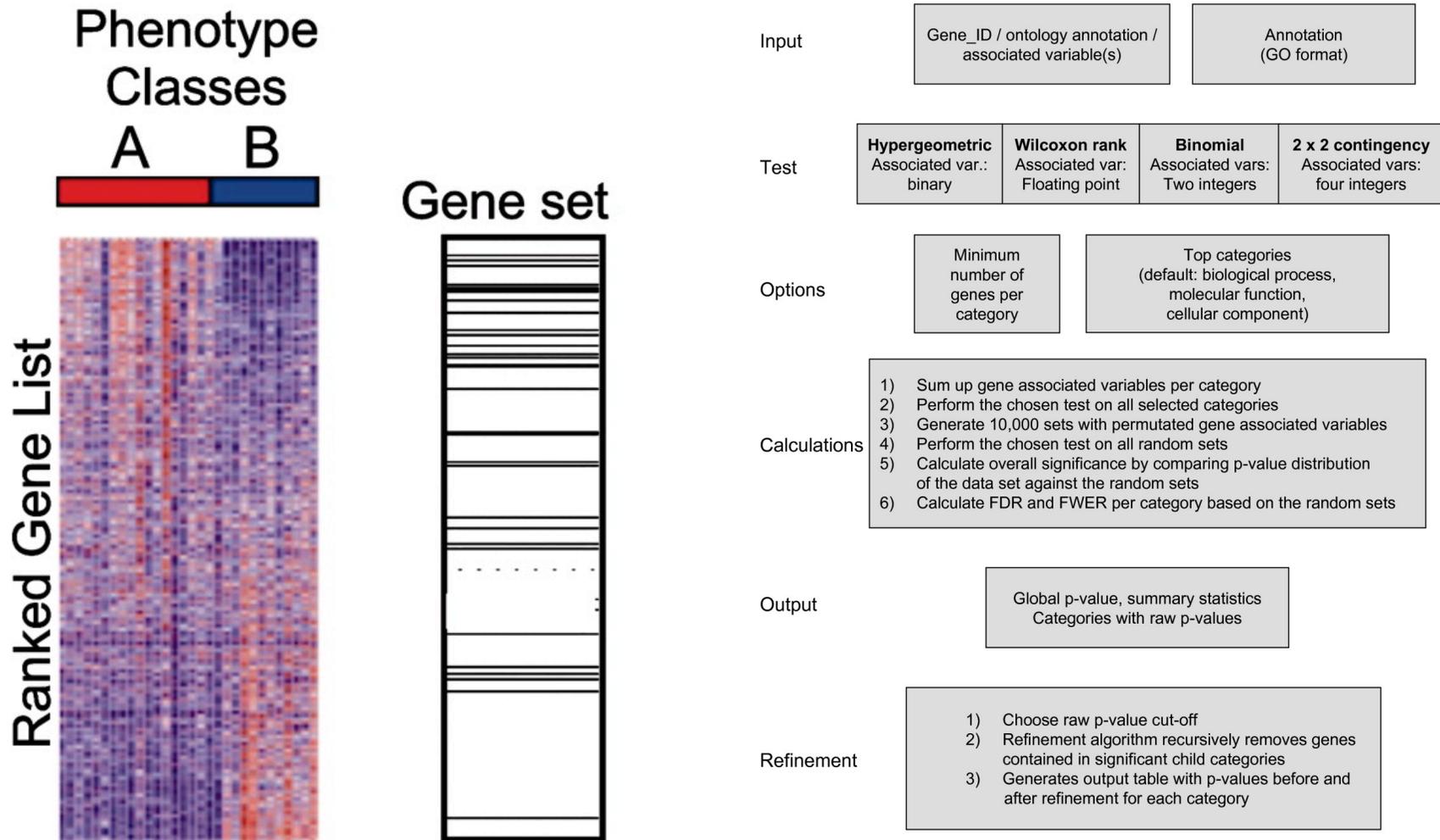
# A simple k=5 split defines ubiquitous genes



# Defining 1,097 LC-enriched genes (14% of 8,011)



# Expression ratios vs. gene functions (e.g., GO)



GSEA: Subramanian et al. (2005), Proc. Natl. Acad. Sci. USA. 102, 15545-15550.  
GO terms via FUNC. Ref.: Prüfer et al. (2007), BMC Bioinformatics 8, 41.

# LC-expressed genes have specific traits

## **Genes expressed more strongly in LC than in whole larvae:**

transcriptional regulation  
cytoskeletal protein binding  
cell adhesion  
intracellular protein transport

# LC-expressed genes have specific traits

## **Genes expressed more strongly in LC than in whole larvae:**

transcriptional regulation  
cytoskeletal protein binding  
cell adhesion  
intracellular protein transport  
cholinergic synaptic transmission  
axon components

# LC-expressed genes have specific traits

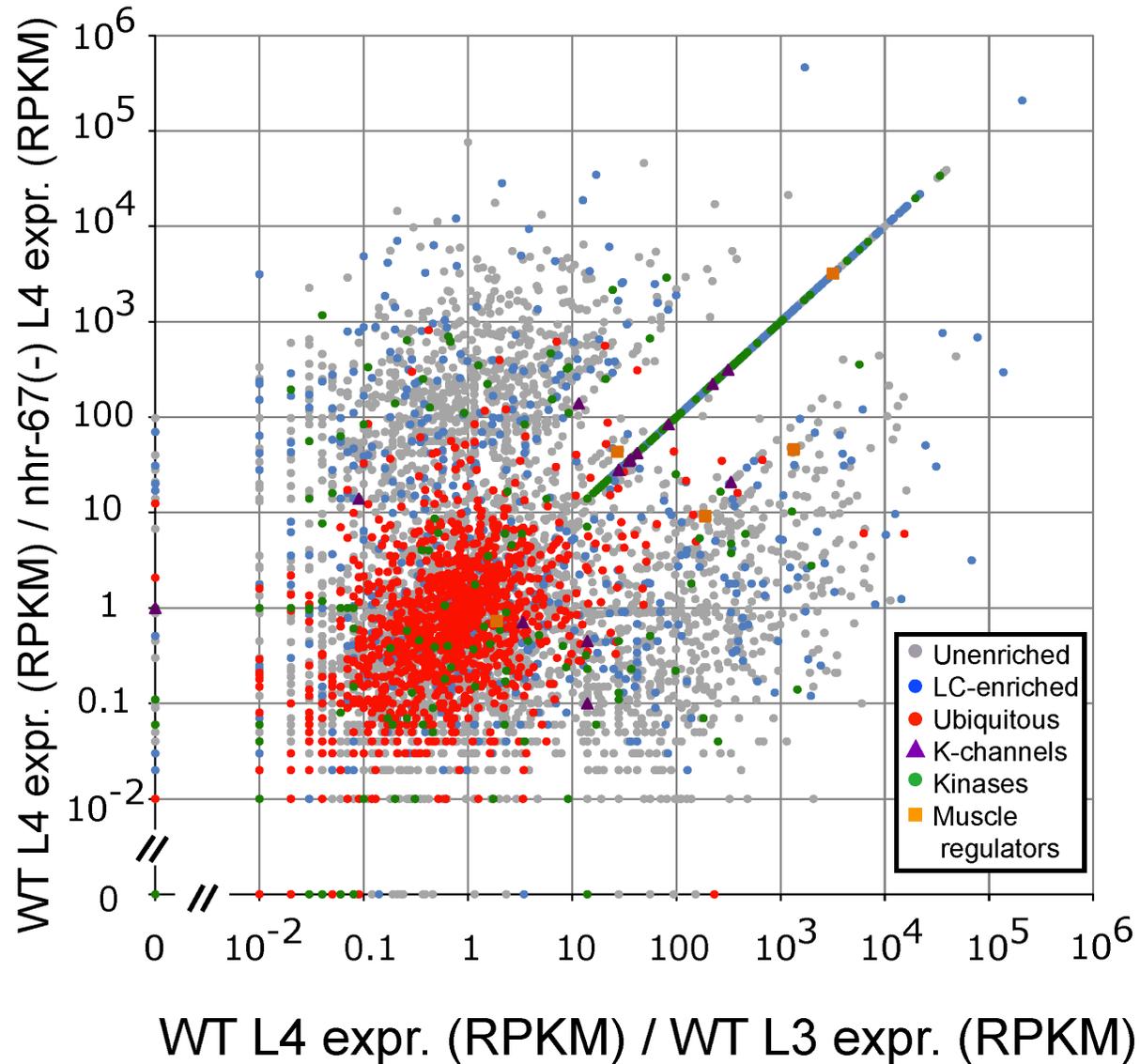
## **Genes expressed more strongly in LC than in whole larvae:**

transcriptional regulation  
cytoskeletal protein binding  
cell adhesion  
intracellular protein transport  
cholinergic synaptic transmission  
axon components

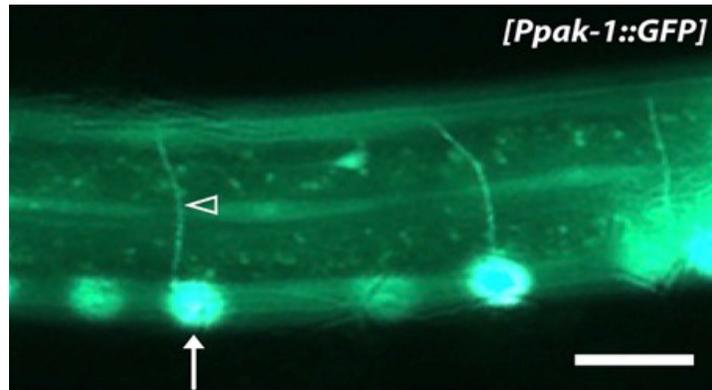
## **Genes upregulated from L3 to L4 by NHR-67:**

potassium channels  
regulators of muscle contraction

# NHR-67-dependent gene upregulation in L4



## "Axon components" include migratory kinases



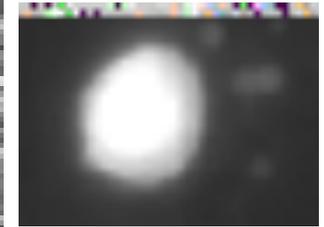
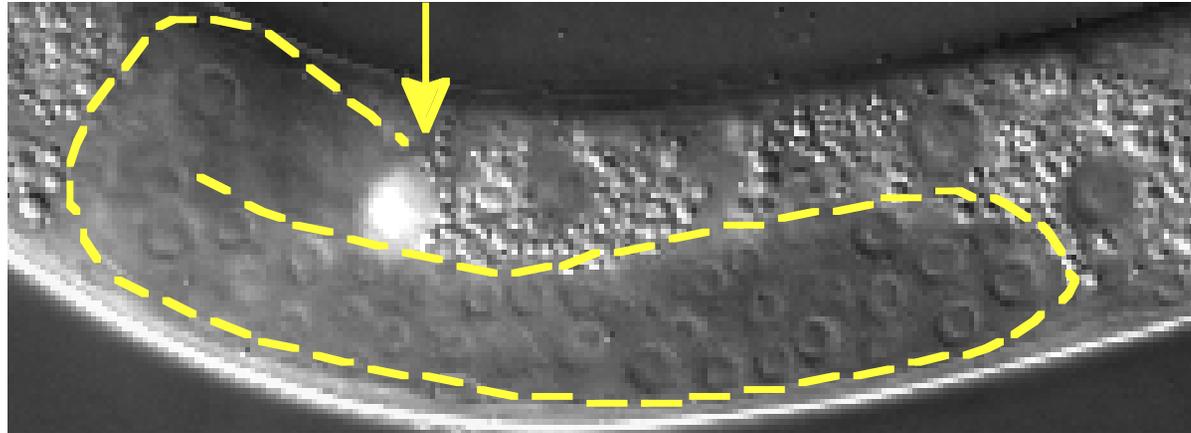
Among the 45 LC genes associated with axon components were *cam-1*, *pak-1*, *sax-1*, *unc-51*, and *vab-1*, which encode protein kinases required for **normal migration or morphology of axons and neurons.**

Another similarity to neurons is that acetylcholine and glutamate (AMPA) receptors are expressed in migrating LCs.

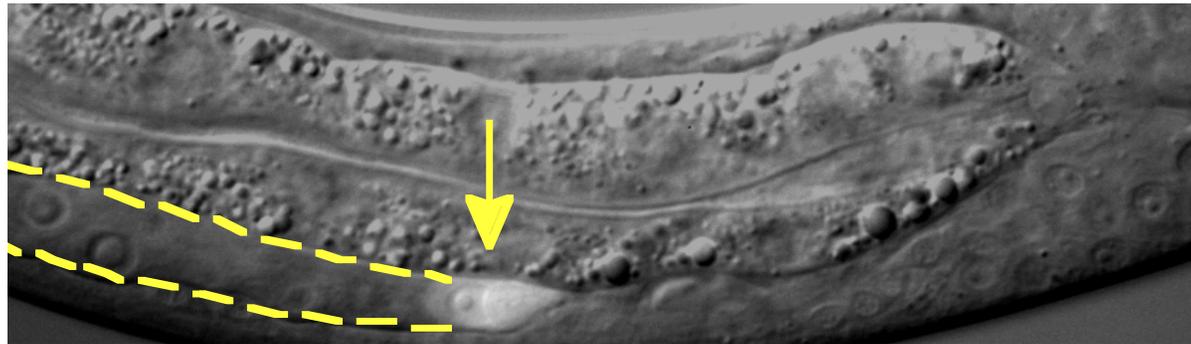
Refs.: Gallegos and Bargmann (2004), *Neuron* 44, 239-249; Hayashi et al. (2009), *Nat. Neurosci.* 12, 981-987; Lucanic et al. (2006), *Development* 133, 4549-4559; Mohamed and Chin-Sang (2006), *Dev. Biol.* 290, 164-176; Ogura et al. (2010), *Development* 137, 1657-1667.

# LCs change shape in the L4 stage

L3  
stage



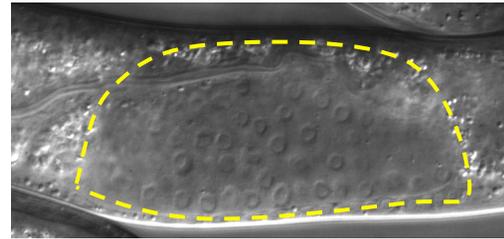
L4  
stage



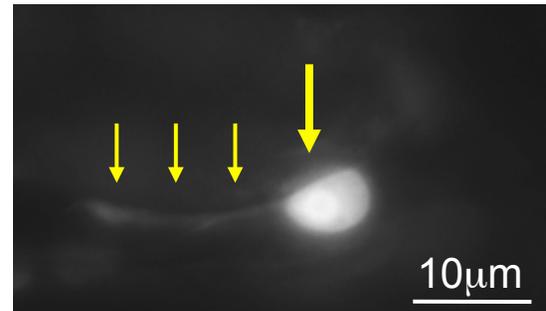
The myosin regulatory light chain genes *mlc-1* and *mlc-2* and the troponin *mup-2* are upregulated by NHR-67 in L4 LCs.

# RNAi phenotypes in 45 of 204 genes (22%)

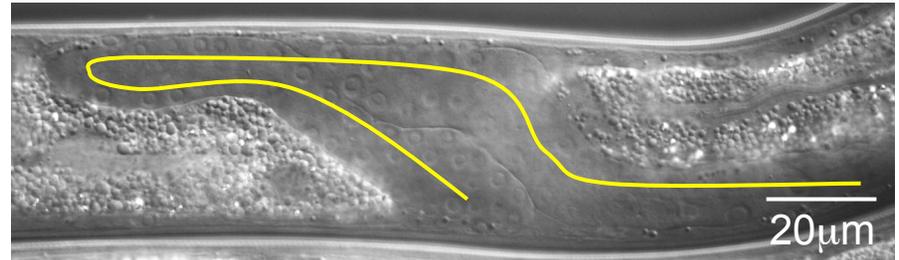
1. **No LC specified:**  
*hlh-2* (E/Da HLH)



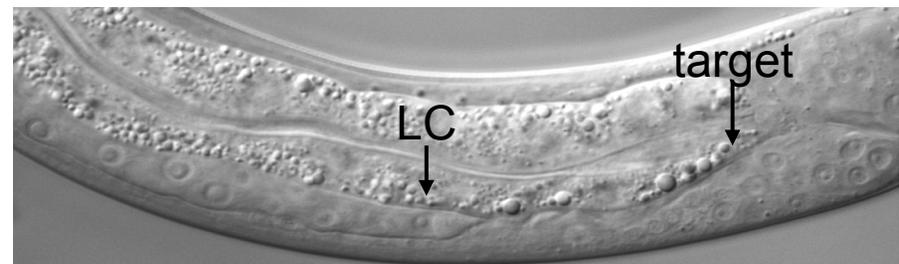
2. **Abnormal LC shape:**  
*inft-1* (INF2 formin)



3. **Abnormal path:**  
*lim-9* (*limpet*/FHL2)



4. **Slow migration:**  
*sphk-1* (sphingosine kinase)



# 14/48 TFs tested are required by the LC

	RNAi phenotypes:				
<b>Genes:</b>	<b>Identity</b>	<b>Shape</b>	<b>Path</b>	<b>Slow</b>	<b>Gonad</b>
<i>hlh-2</i> [E/Da]	---				
<i>daf-12</i> [LXR]		---		---	---
<i>lin-29</i> [ZNF367]	---	---	--	---	

# 14/48 TFs tested are required by the LC

Genes:	RNAi phenotypes:				
	Identity	Shape	Path	Slow	Gonad
<i>hlh-2</i> [E/Da]	---				
<i>daf-12</i> [LXR]		---		---	---
<i>lin-29</i> [ZNF367]	---	---	--	---	
<i>hlh-8</i> [Twist]	-	--		-	
<i>hlh-19</i>	--	-	--	---	-
<i>nhr-97</i>		--	-	---	
<i>egl-5</i> [AbdB]		--		---	---
<i>nob-1</i> [AbdB]		-	---	---	
<i>crh-2</i> [CREB]				---	---
<i>zfp-1</i> [AF10]		--		---	---
ZC123.3 [ZFHX]		-		---	--
<i>pha-1</i>		---	---	--	
<i>gei-13</i>		---	---	---	
T20F7.1 [Zf]		---	---	---	

# 14/48 TFs tested are required by the LC

Genes:	RNAi phenotypes:				
	Identity	Shape	Path	Slow	Gonad
<i>hlh-2</i> [E/Da]	---				
<i>daf-12</i> [LXR]		---		---	---
<i>lin-29</i> [ZNF367]	---	---	--	---	
<b><i>hlh-8</i> [Twist]</b>	-	--		-	

Homologs of TWIST are actively required for cell migration during (EMT-like) cancer cell metastasis in mammals, and during *Drosophila* gastrulation.

<i>crh-2</i> [CREB]				---	---
<i>zfp-1</i> [AF10]		--		---	---
ZC123.3 [ZFHX]		-		---	--
<i>pha-1</i>		---	---	--	
<i>nei-13</i>		---	---	---	

Refs.: Yang et al. (2004), Cell 117, 927-939; Kölsch et al. (2007), Science 315, 384-386.

# Some other known migration genes rediscovered

*crh-2* =~ *Drosophila* SLBO, C/EBP $\gamma$  TF  
required for border cell migration

*sphk-1* == sphingosine kinase;  
sphingosine-1-phosphate is generally implicated in migration

*lim-9* == mammalian FHL2 inhibits sphingosine kinase, and  
is involved in colon cancer invasion and dendritic cell migration

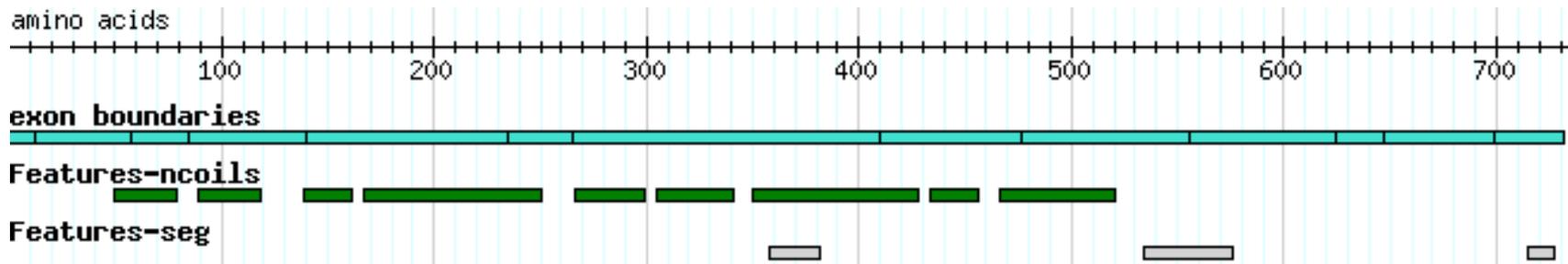
Refs.: Borghese et al. (2006), *Dev. Cell* 10, 497-508; Wang et al. (2006), *Dev. Cell* 10, 483-495; Hayashi et al. (2009), *Arterioscler. Thromb. Vasc. Biol.* 29, 909-914; Fyrst et al. (2010), *Nat. Chem. Biol.* 6, 489-497; König et al. (2010), *J. Immunol.* 185, 1466-1475; Zhang et al. (2010), *Carcinogenesis* 31, 1220-1229.

# However, 'conserved unknowns' also found

*tpra-1* == TPRA40 (7TM) inhibits embryonic cell division

*maea-1* == MAEA/EMP receptor is required for developing erythroblasts to bind macrophages and mature into erythrocytes;  
*may* also have migratory phenotype in cell culture

*ccc-1* ('conserved coiled-coil protein') ==  
C10orf118 has no known function whatsoever



Refs.: Aki et al. (2008), J. Cell. Physiol. 217, 194-206;

Chasis et al. (2008), Blood 112, 470-478; Manjit Hanspal, pers. comm., 25 Feb 2013.

# Potassium channels are, indeed, required for normal LC migration

	RNAi phenotypes:			
Genes:	Identity	Shape	Path	Slow
<i>twk-13</i>	-	--	---	---
<i>twk-18</i>			---	-

There are 46 *twk* genes in *C. elegans*, of which 13 are expressed in LCs.

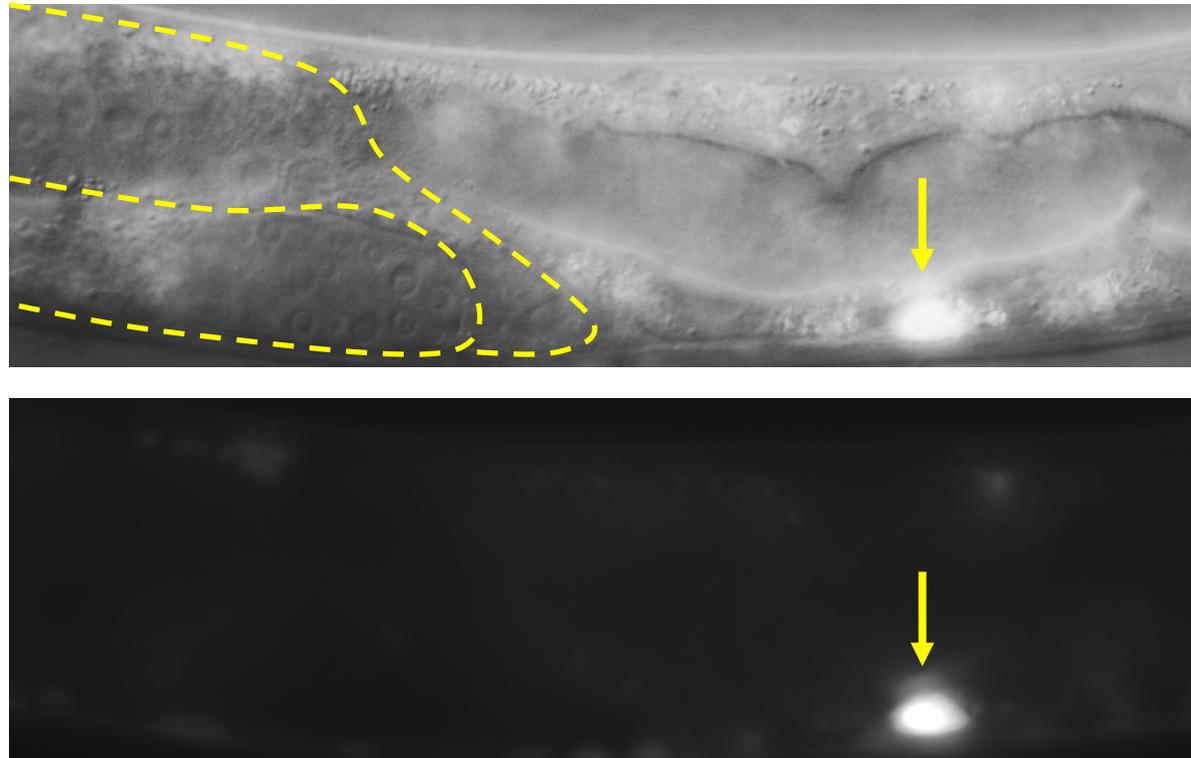
Both *twk-13*(RNAi) and *twk-18*(RNAi) LCs have delayed return from the dorsal to the ventral body wall, a subset of the phenotypes in *nhr-67*(RNAi) LCs.

TWK potassium channels have had no known role in migration, but there is evidence, for a few TWKs, that they can be mechanosensory.

Ref.: Mathie et al. (2010), J. Physiol. 588, 3149-3156.

# Cohesin and condensin subunits enable LC-gonad attachment

*him-1*/SMC1  
*mix-1*/SMC2  
*smc-3*/SMC3  
*dpy-27*/SMC4  
*smc-4*/SMC4



Cohesins can regulate genes as well as join chromosomes;  
condensins may as well, though this is less studied.

Refs.: Seitan et al. (2006), PLoS Biol. 4, e242; Wood et al. (2010), Nat. Rev. Genet. 11, 391-404; Pauli et al. (2010), Curr Biol., 10.1016/j.cub.2010.09.057.

# Major sperm proteins expressed in L4-stage LCs

## MSP-domain protein family 1

	L3	L4	<i>nhr-67</i> (-)	Larvae
D2007.2	5.25	0	0	0.51
F42A9.7	0	1.3	0	11.14
C35E7.9	0	1.59	0	13.77
C35D10.1	0	1.92	0	2.62
F21H7.5	0	2.27	0	17.1
T13F2.12	0	3.41	0	0
ssp-32	0	7.97	0	0.24
F55C5.1	0	8.2	13.06	0.84
W03B1.5	0	10.94	0	1.27
ssp-10	0	14.46	4.15	5.54
ssp-31	0	34.1	0	0.22
C34D4.3	0	38.94	0.54	5.15
ssp-16	0	39.52	3.23	2.64
ssp-19	0	88.92	0	3.31
ssp-11	0	481.57	1.11	26.44
ssp-9	0	774.75	1.13	38.25

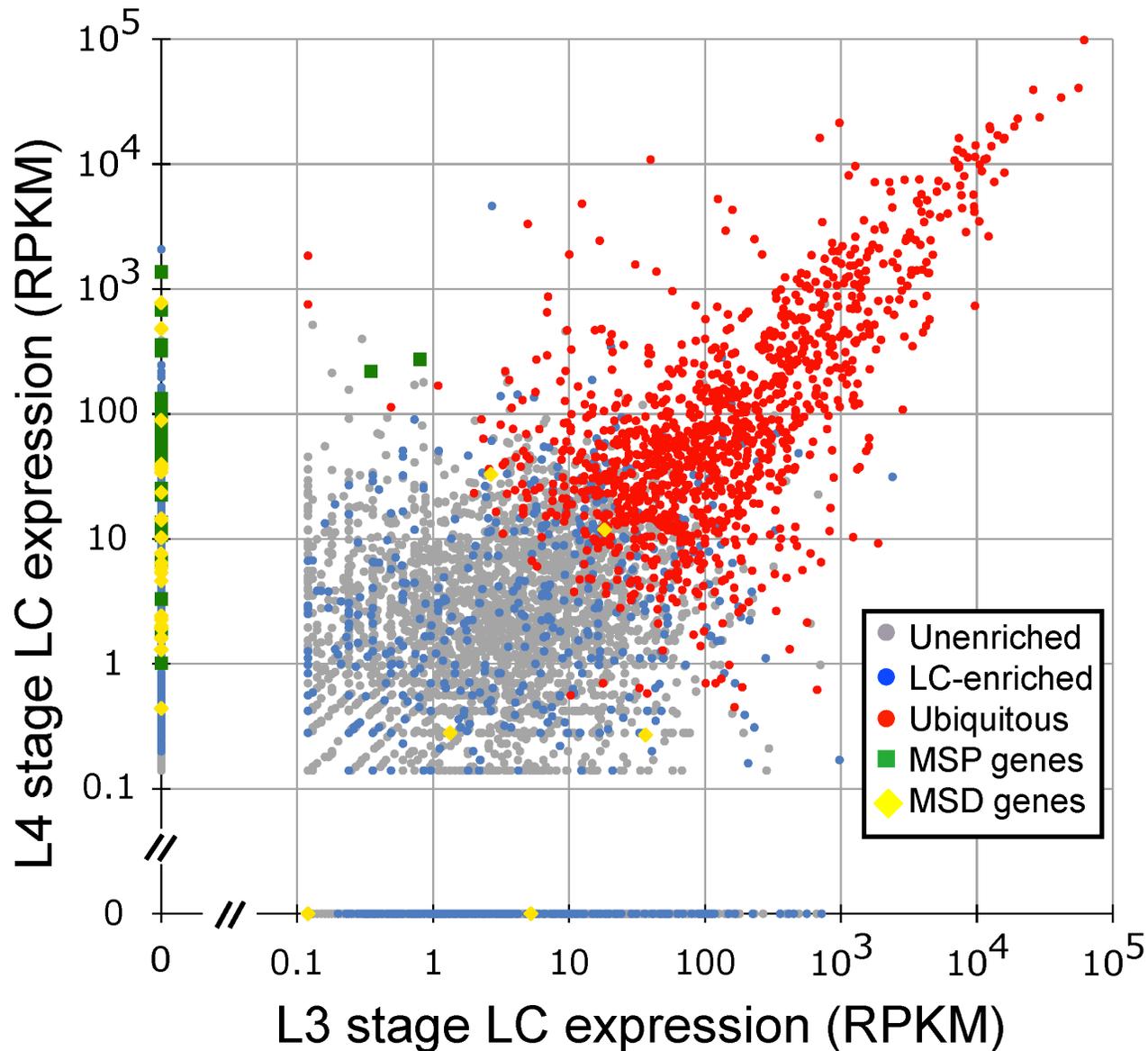
## MSP-domain protein family 2

	L3	L4	<i>nhr-67</i> (-)	Larvae
msp-113	0	22.47	2.27	7.38
ZK546.3	0	23.53	0.58	4.23
msp-142	0	25.31	7.31	6.59
Y59E9AR.	0	34.27	7.15	16.43
Y59E9AR.	0	36.62	7.06	16.02
msp-31	0	46.04	1.53	12.69
msp-81	0	47.1	0	22.41
msp-78	0	53.04	0	9.62
msp-51	0	54.22	3.61	21.6
msp-36	0	63.18	0	13.48
msp-76	0	63.98	24.73	29.09
msp-55	0	73.16	1.54	20.13
msp-38	0	77.84	0	20.46
msp-50	0	84.66	0	38
msp-64	0	90.54	0	5.48
msp-57	0	105.35	1.03	63.28
msp-33	0	109.43	0.51	18.84
msp-59	0	119.63	2.05	64.57
msp-65	0	133.43	1.02	49.23
msp-53	0.35	219.67	2.28	39.48
msp-19	0.8	273.78	22.52	29.01
msp-49	0	320.51	0	20.03
msp-45	0	358.54	0.47	17.23
msp-40	0	681.52	215.36	27.99
msp-3	0	1372.22	4.64	17.97

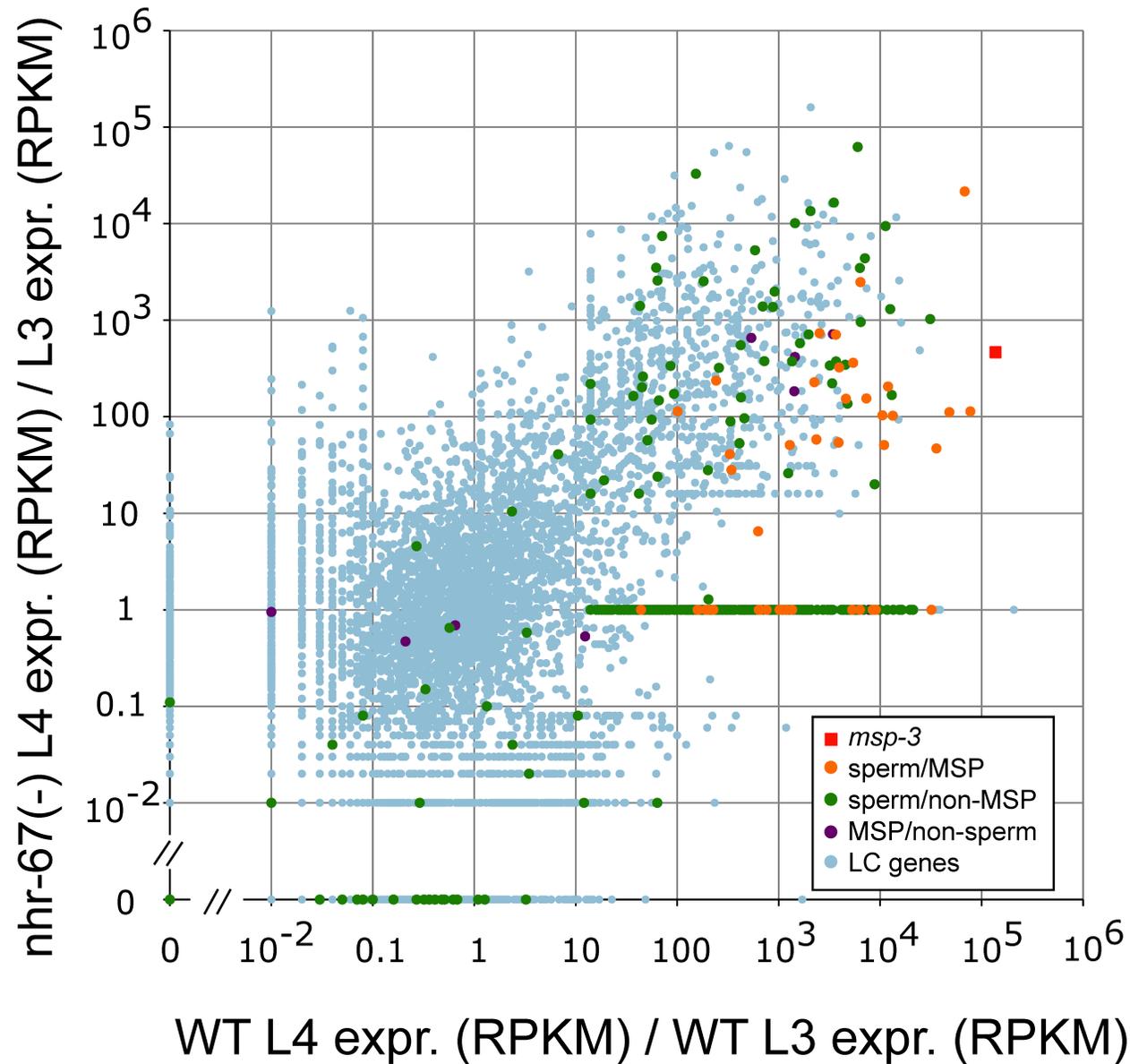
## MSP-domain protein family 2

msp-63	0	1.01	1.13	0.84
msp-79	0	6.59	0	4.14
msp-56	0	10.73	0	4.03
msp-77	0	12.01	0	7.19
msp-152	0	12.86	0.51	8.84
msp-10	0	13.57	0	6.79
Y59H11A1	0	14.25	1.83	5.21

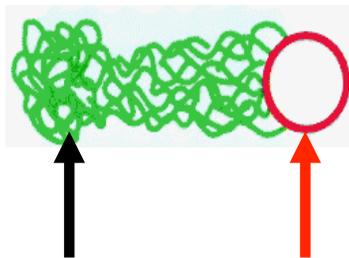
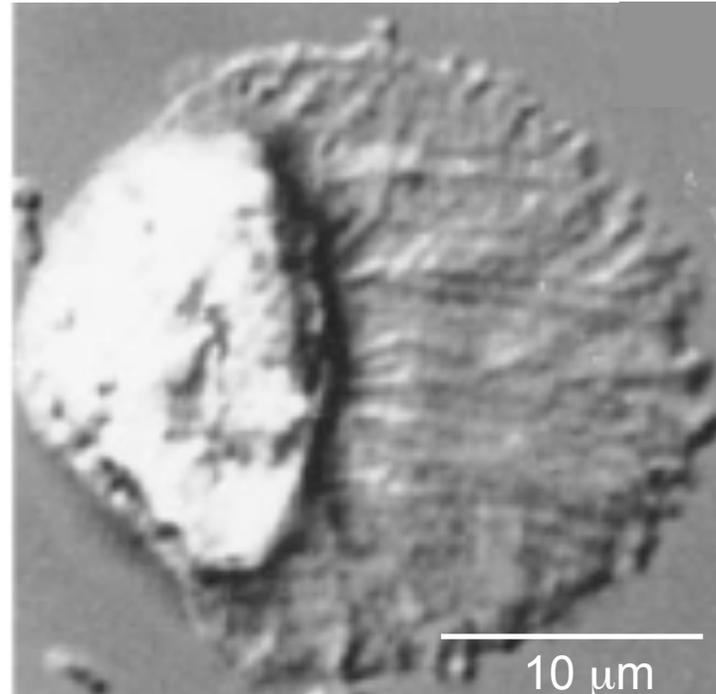
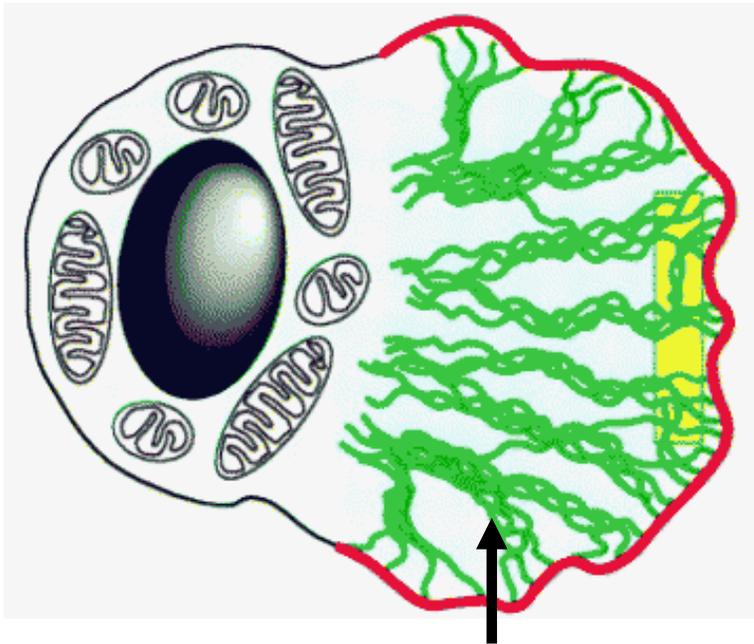
# MSP and MSD genes are heavily L4-specific



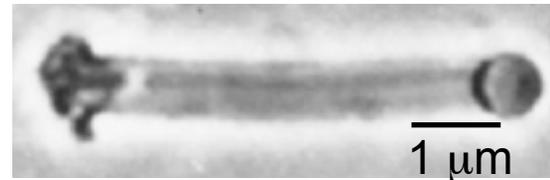
# MSP and MSD genes are NHR-67-dependent



# MSP, not actin, drives nematode sperm motility

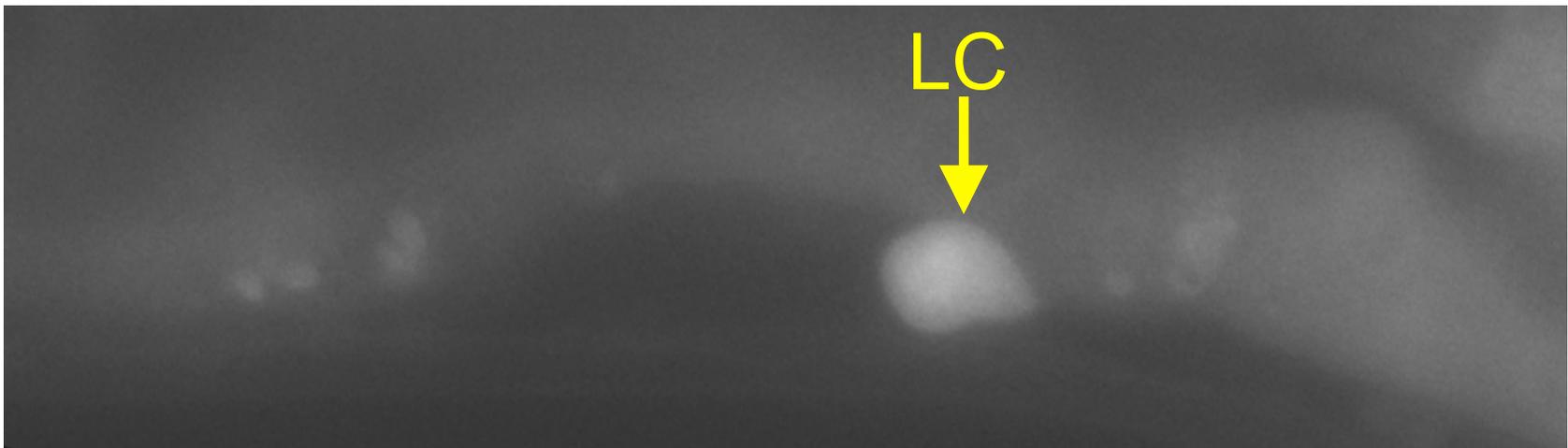
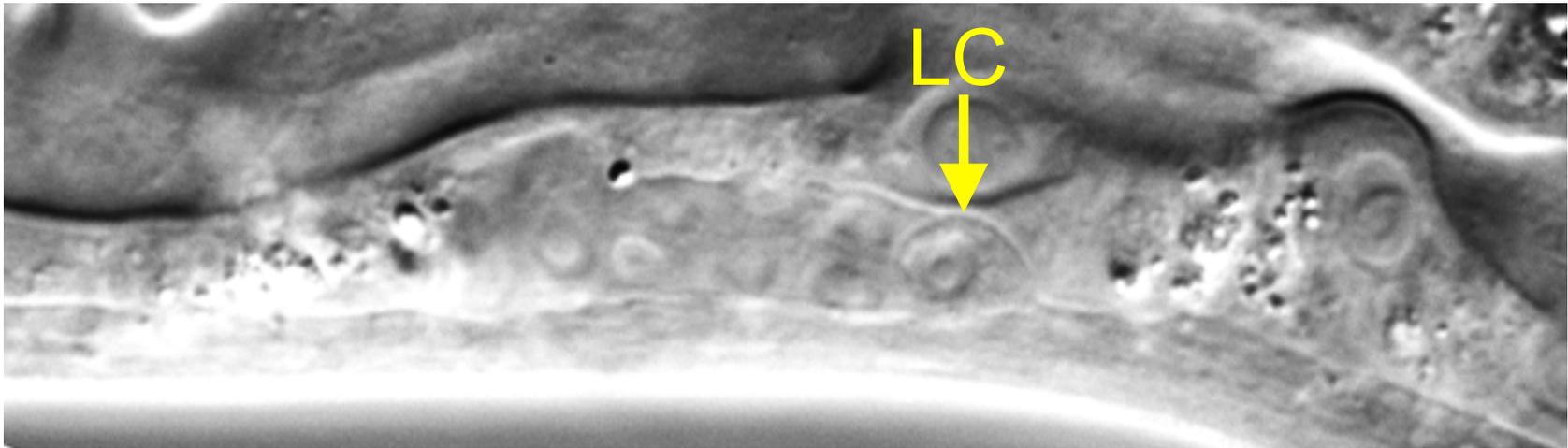


MSP fibers and membrane vesicle



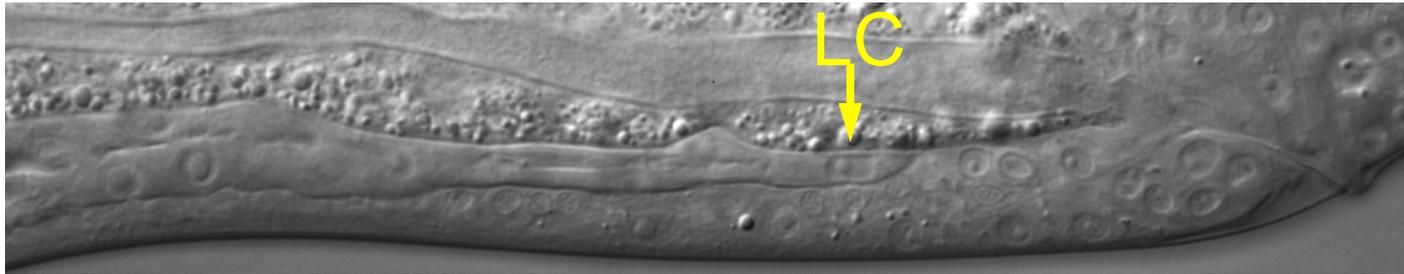
Refs.: Italiano et al. (1996), *Cell*, 84, 105-114; Theriot (1996), *Cell*, 86, 1-4.

MSP::YFP is expressed in L4-stage linker cells

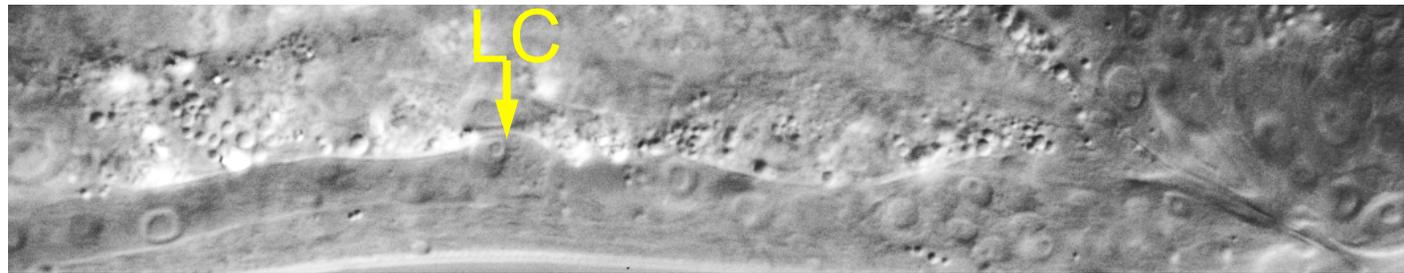


# *msp-3*(RNAi) LCs are slow and abnormally round

wild  
type



*msp-3*  
RNAi



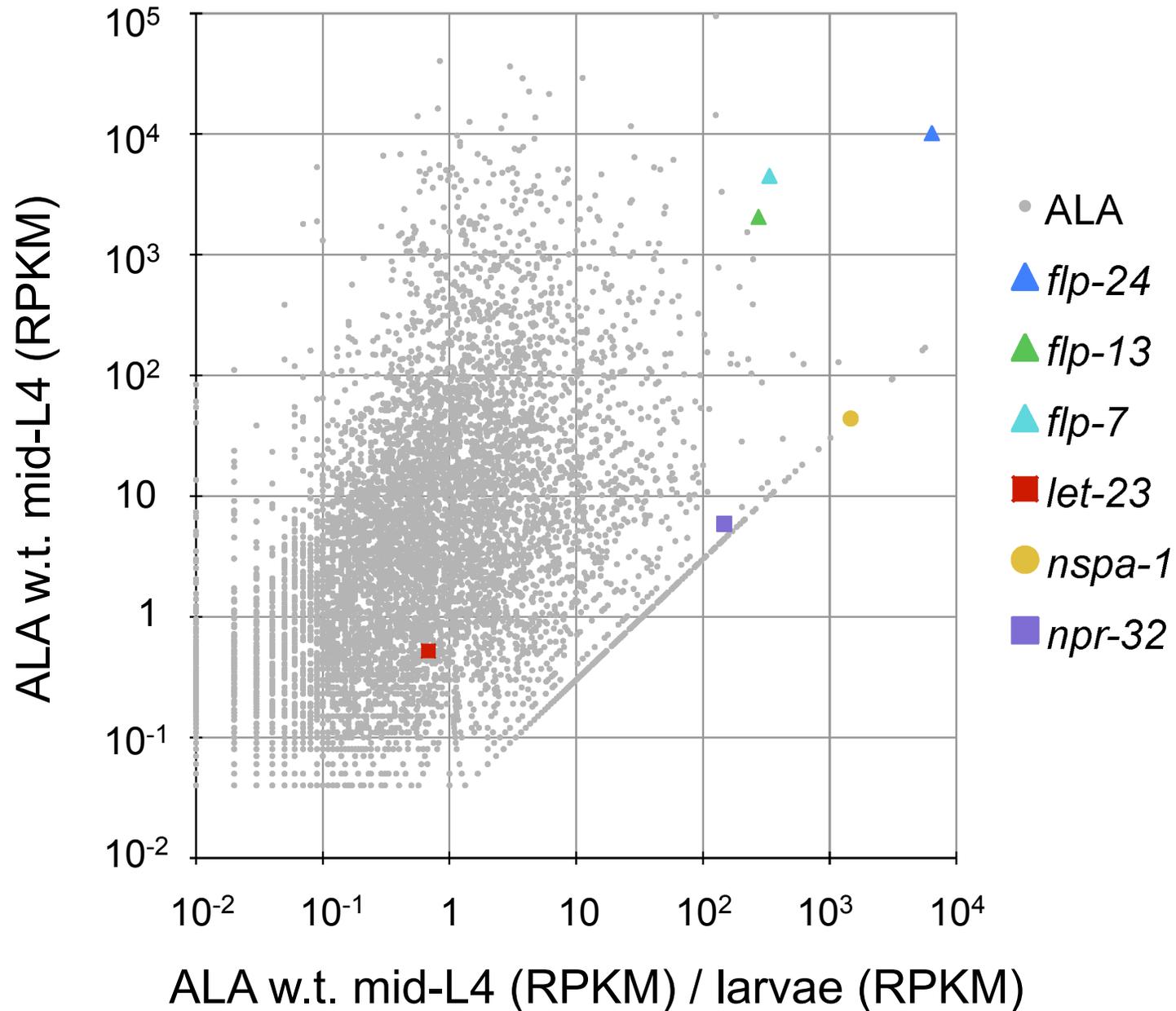
## Conclusions from linker cell RNA-seq

1. TWK potassium channels might have mechanosensory role in cell migration.
2. MSPs can no longer be considered only 'sperm' proteins, and thus should be considered as possible motor proteins in other cell types or organisms.
3. Other novel conserved proteins may also act during migration of animal cells, e.g., MAEA.

## More general conclusions

1. Highly dynamic single cells can be transcriptionally profiled.
2. Gene sets are large but manageable.
3. Profiling is surprise-driven rather than hypothesis-driven.

# 8,133 genes in neuroendocrine ALA neurons



# Thanks:

Mihoko Kato

Steven Kuntz

Miriam Goodman

Yen-Ping Hsueh

Elly Chow

Ali Mortazavi

Brian Williams

Igor Antoshechkin

Jacobs Genome Center

Linker cell dissections, RNAi, and GFP

Dissection of *ceh-13/lin-39* Hox elements

AFD/ASER/PLML dissections

AWC RNA-seq

ALA RNA-seq

Early RNA-seq analyses

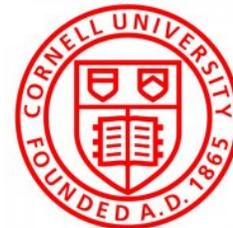
cDNA library construction

Optimized sequencing protocols

Illumina sequencing



HHMI





# Profile of a chemosensory neuron (AWC)

